REMARKS

Reconsideration and entry of the present amendment after final is respectfully requested.

Status of the Claims:

Claims 2-8, 12, 14, and 18-38 were pending after submission of 13 June 2003. Claims 9-11 have been cancelled in the current amendment, they were previously withdrawn from examination. Applicants reserve the right to pursue the contents of these claims in continuing applications. Claims 2-8, 12, 14, and 18-38 remain under examination after amendment. Claims 12, 14, and 20 have been amended regarding the encoded polypeptide, the phrase "involved in DNA double strand break repair" has been replaced with the phrase "binds to a MRE11 polypeptide". Support for the amendments is found in the claims as originally filed, and throughout the specification, particularly page 1, lines 30-32. No new matter has been added.

Rejections under 35 U.S.C. §112, 1st Paragraph - Enablement:

Claims 2-8, 12, 14, and 18-38 are rejected under 35 U.S.C. §112, first paragraph as failing to comply with the enablement requirement. The rejection is repeated for the reasons of record as set forth in the Office Action of 1/13/03.

The Action asserts, briefly, that given the complexity of DNA repair, including the involvement of other proteins, it is unclear what agronomic benefit a transgenic plant with modulated Rad50 levels would have, and unclear how modulating the level of Rad50 would affect DNA repair or polynucleotide integration. Therefore the Action concludes that one skilled in the art would not know how to use SEQ ID NO: 1, and sequences having 90% and 95% to the disclosed sequences, without undue experimentation.

Applicants respectfully disagree, for the reasons of record (e.g., see response filed 6/13/03), some of which may be reiterated for clarity. Applicants note that

claims 12, 14, and 20 have been amended, as described above, to recite polypeptides that "bind to a MRE11 polypeptide".

Applicants have provided full-length sequences, demonstrated conserved functional motifs (e.g., page 2, lines 17-24, and Example 4), extensive overall homology, direction regarding nucleotide and amino acid substitutions, and percent identity comparisons (e.g., Appendix A and D; page 5, line 20 – page 6; and line 25; page 16, line 1 – page 20, line 20 of the specification), Rad50 function in DNA break repair and interaction with Mre11 (e.g., page 1, line 15 – page 2, line 14), vector construction, plant transformation, and methods to modulate the level of Rad50 (e.g., page 37, line 5 – page 42, line 22; page 44, line 28 – page 47, line 25; page 47, line 28 – page 49, line 26; page 50, line 29 – page 52, line 31; and page 53, line 1 – page 54, line 6). The Examiner is reminded that the need for routine experimentation and screening is not considered undue (see MPEP 2164.01):

The test of enablement is not whether any experimentation is necessary, but whether, if experimentation is necessary, it is undue. In re Angstadt, 190 USPQ 214 (CCPA 1976). An extended period of experimentation may not be undue if the skilled artisan is given sufficient direction or guidance. In re Colianni, 195 USPQ 150 (CCPA 1977) (Miller, J., concurring). The experimentation required, in addition to not being undue, must not require ingenuity beyond that expected of one of ordinary skill in the art. In re Angstadt, supra. For example, in one instance a "few hours" of experimentation to determine process parameters was not considered to be undue in view of the nature of the invention (preparation of oxygenated hydrocarbons). In re Borkowski, 164 USPQ 642 (CCPA 1970). In Tabuchi v. Nubel, 194 USPQ 521 (CCPA 1977) a screening procedure which took 15 calendar days was not considered undue experimentation because the test was both simple and straightforward and because of its demonstrated success in producing the desired result.

Further, the Examiner has not given specific reasons, or provided specific citations to support the conclusions that modulation of the level of Rad50 would not have an effect on the plant.

Modulation of Rad50, thereby modulating the level of DNA repair in the plant, is predicted to increase the efficiency of incorporation of heterologous nucleic acids

in the genome of a plant. For the record, Applicants disagree with the Examiner's conclusion that modulation of Rad50 must result in an agronomic benefit to the transgenic plant. Applicants utility, to use Rad50 to modulate the efficiency of incorporation of a transgene into the plant genome (page 2, lines 10-14) does not require that Rad50 confer any particular agronomic phenotype to the plant.

Applicants note that they have provided another component of the DNA repair complex, Mre11, in U.S. Patent 6,646,182, issued Nov. 11, 2003, thereby demonstrating the presence of Mre11 in plants. Also, Gallego *et al.* have demonstrated Rad50 involvement in DNA repair and meiosis in *Arabidopsis* (*Plant J.* 25:31-41 2001, submitted in Appendix C). Applicants have modified the Multiple Sequence Alignment (Appendix A) to include the conserved motifs shown in Example 4 of the specification, currently presented as Appendix D. Applicants note that the alignment is similar to the one shown by Gallego *et al.* (supra), which shows higher conservation of the N- and C-terminal regions of Rad50. Appendix D also includes the results of an analysis performed using the Lion BioScout software package, and Hmmerpfam search using GCG software, both of which detect the known Rad50 Zn-hook Pfam domain (e = 6.9e-07).

Applicants assert this support fully enables one of skill in the art to make and use the full-breadth of the invention without undue experimentation. Absent of any evidence from the Examiner supporting the conclusion that modulation of Rad50 would not effect the plant, Applicants believe this rejection cannot be supported. Therefore Applicants respectfully request reconsideration and that the rejection of claims 2-8, 12, 14, and 18-38 under 35 U.S.C. §112, first paragraph for lack of enablement be withdrawn.

Rejections under 35 U.S.C. §112, 1st Paragraph, Written Description:

Claims 2-8, 12, 14, 20 and 23-38 are rejected under 35 U.S.C. §112, first paragraph, as containing subject matter not sufficiently described in the specification

to indicate the inventor(s) had possession of the invention. The rejection is repeated for the reasons of record as set forth in the Office Action of 1/13/03.

Briefly, the Action asserts that the Applicant has not described a representative sample of the genus, and that the conserved domains are not unique to Rad50 proteins, but are common to all DNA repair proteins. The Action also asserts that Example 14 of the Revised Interim Written Description Guidelines is not applicable because claims drawn to polynucleotides having 95% sequence identity are not rejected in this Action.

Applicants respectfully disagree, for the reasons of record (e.g., see response filed 6/13/03), some of which may be reiterated for clarity. As is stated in the MPEP 2163 (see p. 2100-168) the written description for a claimed genus may be satisfied by "disclosure of relevant, identifying characteristics, i.e., structure or other physical and/or chemical properties, by functional characteristics coupled with known or disclosed correlation between function and structure, or by a combination of such identifying characteristics, sufficient to show that the applicant was in possession of the claimed genus." Applicants respectfully submit that the present application meets this standard by the disclosure of the structures of the full-length Rad50 polynucleotide (SEQ ID NO: 1), which encodes a full-length Rad50 polypeptide (SEQ ID NO: 2) which have the structural/chemical properties of significant sequence identity to known Rad50 polynucleotides and polypeptides, conserved domains (see, for example, Example 4 and the Multiple Sequence Alignment), and the functional characteristic of Rad50 polypeptide binding to MRE11 polypeptides.

Regarding the applicability of Example 14, for the record Applicant notes that while claims 18 and 21, directed to sequences having 95% identity to either SEQ ID NO: 1 or SEQ ID NO: 2, are not included in this rejection, they are also not allowed. Also, the Examiner has not made clear why Example 14 does not apply, other than focusing on the specific percent identity recited and not the substantive features of the structural and functional elements. Applicants maintain that the current claims, including those directed to 90% sequence identity, meet the Written Description

standard as set forth in the 35 U.S.C. §112, described MPEP, and shown in Example 14 of the Revised Interim Written Description Guidelines.

Applicants have coupled structural, chemical, and functional properties to describe polynucleotides having 90% and 95% sequence identity to SEQ ID NO: 1, or encoding polypeptides that have 90% and 95% sequence identity to SEQ ID NO: 2 such that a person skilled in the art can envisage the claimed invention, thereby meeting the 35 U.S.C. §112, first paragraph written description requirement. Therefore Applicants respectfully request reconsideration and that the rejection of claims 2-8, 12, 14, 20 and 23-38 under 35 U.S.C. §112, first paragraph for lack of written description be withdrawn.

CONCLUSION

In light of the foregoing remarks and amendments, it is believed that claims 2-8, 12, 14 and 18-38 are in condition for allowance. Withdrawal of the outstanding rejections and allowance of all of the remaining claims is respectfully requested. The Examiner is invited to telephone the Applicants representative to expedite prosecution and allowance.

Respectfully submitted,

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APPENDIX C

The Plant Journal (2001) 25(1), 31-41

Disruption of the *Arabidopsis RAD50* gene leads to plant sterility and MMS sensitivity

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Summary

The Rad50 protein is involved in the cellular response to DNA-double strand breaks (DSBs), including the detection of damage, activation of cell-cycle checkpoints, and DSB repair via recombination. It is essential for meiosis in yeast, is involved in telomere maintenance, and is essential for cellular viability in mice. Here we present the isolation, sequence and characterization of the Arabidopsis thaliana RAD50 homologue (AtRAD50) and an Arabidopsis mutant of this gene. A single copy of this gene is present in the Arabidopsis genome, located on chromosome (I. Northern analysis shows a single 4.3 Kb mRNA species in all plant tissues tested, which is strongly enriched in flowers and other tissues with many dividing cells. The predicted protein presents strong conservation with the other known Rad50 homologues of the amino- and carboxy-terminal regions. Mutant plants present a sterility phenotype which co-segregates with the T-DNA insertion. Molecular analysis of the mutant plants shows that the sterility phenotype is present only in the plants homozygous for the T-DNA insertion. An in vitro mutant cell line, derived from the mutant plant, shows a clear hypersensitivity to the DNA-damaging agent methylmethane sulphonate, suggesting a role of RAD50 in double-strand break repair in plant cells. This is the first report of a plant mutated in a protein of the Rad50-Mre11-Xrs2 complex, as well as the first data suggesting the involvement of the Rad50 homologue protein in meiosis and DNA repair in plants.

Keywords: Arabidopsis, RADSO, DSB repair, sterility, recombination.

Introduction

The repair of DNA double-strand breaks (DSBs) involves genetic recombination. Fundamentally, two different forms of recombination are involved: those involving DNA sequence homology between the participating DNA molecules, and those that appear to act independently of such homology, called homologous recombination (HR) and non-homologous end-joining (NHEJ), respectively. Considerable advances in the understanding of these mechanisms have been made in recent years, both in terms of the proteins involved and of their relative contribution to repair of DSBs in different organisms and/or cell types. These issues are the subject of a number of recent reviews (Fox and Smith, 1998; Jeggo, 1998; Paques and Haber, 1999; Petrini etal., 1997; Roeder, 1997; Smith, 1998; Smith and Nicolas, 1998; Tsukamoto and lkeda, 1998).

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Much of our understanding of the proteins implicated in the processes of recombination and DSB repair comes from studies of the yeast Saccharomyces cerevisiae. Homologous recombination in yeast cells is largely under the control of the genes in the RAD52 epistasis group, which include RAD50-59, XRS2 and MRE11 (Ajimura etal., 1993; Game, 1993; Haynes and Kunz, 1981). These genes were mostly identified as being needed for the repair of ionizing radiation-induced DNA damage, and are also needed for melosis. Deletion of these genes in yeast generates defects in both recombination and DNA repair with differing phenotypes in terms of recombination and the repair of DNA double-strand breaks. Mutants of rad51, rad52, rad54, rad55, rad57 and rad59 show defects in HR, while rad50 and mra11 mutants have shown defects in NHEJ and are hyper-rec for HR.

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Double-strand breaks are repaired by both HR and NHEJ in all cell types examined; however, the preferred mechanism of recombinational DNA repair differs significantly in a given organism. In particular, it is clear that nonhomologous recombination is much more frequent than homologous recombination in mammalian and plant cells, whereas yeast cells rely almost entirely on homologybased recombinational DNA repair. The basis of the strong preference for illegitimate recombination in mammalian and plant cells is not fully understood. Both pathways have a common initial substrate (OSB), the processing of which may be channelled into either HR or NHEJ. This channelling is apparently determined by the initial metabolism of the DSB itself, and it has recently been suggested that it may result from a competition for binding the DNA ends between the Ku and Rad52 proteins (VanDyck et al., 1999). In this context, the Rad50/Mre11/Xrs2 complex is of particular interest, as mutants show a weak hyper-HR and strong hypo-NHEJ phenotype in mitotic yeast cells (Boulton and Jackson, 1998; Moore and Haber, 1996; Schiestl et al., 1994; Tsukamoto et al., 1996; Tsukamoto et al., 1997). However, in some assays RAD50 is needed for intrachromosomal recombination (Elias-Arnanz et al., 1996; Rattray and Symington, 1995; Tran et al., 1995). Rad50 mutants show altered processing of DNA ends during recombination (Ivanov et al., 1994; Sugawara and Haber, 1992). This complex has also been implicated in the regulation of the response of yeast cells to DNA damage (Kiromani and Muniyappa, 1997; Lee et al., 1998).

Homologues of many of these yeast genes have been cloned from different organisms (see reviews by Kanaar and Hoeijmakers, 1997; Paques and Haber, 1999), including RAD50 genes from S. cerevisiae (Alani et al., 1989); S. pombe (Saunders, 1999); man (Dolganov et al., 1996); mouse (Kim et al., 1996); and Caenorhabditis elegans (Offenberg and Heyting, 1996). Rad50 shows homology to the Escherichia coli SbcC protein (Sharples and Leach, 1995), and has been shown to form a complex with the Mre11, Xrs2 and Lig4 proteins. The Rad50/Mre11 complex is involved in DSB repair via NHEJ in mammalian cells, and the null mouse rad50 mutant is inviable both in cultured ES cells and in developing mouse embryos (Luo et al., 1999). This cell-lethal phenotype has also been seen in mouse mre11 mutants (Xiao and Weaver, 1997), as well as chicken (Sonoda et al., 1998) and mouse rad51 mutants (Lim and Hasty, 1996; Sharan et al., 1997; Tsuzuki et al., 1996). Interestingly, rad52 mutant mice are viable and show increased radioresistance (Rijkers et al., 1998; Yamaguchi-Iwai et al., 1998).

A number of radiation-sensitive plant mutants have been isolated in recent years (reviewed by Britt, 1999; Gorbunova and Levy, 1999; Mengiste and Paszkowski, 1999). The mutated genes of methylmethane sulphonate (MMS) and UV-hypersensitive Arabidopsis mutants have recently been identified: a ribosomal S27 protein (Revenkova et al., 1999); the Arabidopsis Rad1 homologue protein (Gallego et al., 2000; Liu et al., 2000); and a member of the structural maintenance of chromosomes protein family (SMC; Mengiste et al., 1999; Strunnikov, 1998; Strunnikov et al., 1993). Rad50 is also a member of this family. Plant homologues of RAD51 (Doutriaux et al., 1998; Smith et al., 1996) and MRE11 (Hartung and Puchta, 1999) have been isolated, and the rapid progress of the Arabidopsis sequencing project makes it likely that many other genes implicated in these processes will be isolated in the near future.

As part of our investigation of the control of the early events of DSB repair and recombination in plants, we have isolated the *Arabidopsis thaliana* homologue of the *RAD50* gene, and here we describe the isolation, sequencing and preliminary characterization of this gene. Furthermore, we have identified a rad50 mutant plant that presents a sterility phenotype in agreement with the role of this protein in meiosis in yeast cells. We also show that homozygous mutant cell lines present hypersensitivity to MMS, suggesting (as has been shown in yeast cells) a role for *RAD50* in DSB repair in plants. This is the first evidence of the implication of the *RAD50* gene in meiosis as well as in DSB repair in plants.

Results and Discussion

The Arabidopsis thaliana RAD50 homologue

A 700 bp A. thaliana cDNA clone with homology to the carboxy-terminal region of the yeast Rad50 gene was detected in a screen of the Arabidopsis cDNA expression library with an antibody against a peptide sequence from the cytoplasmic domain of vertebrate beta 1 integrin. Based on this sequence (a kind gift from P. Nagpal and R. Quatrano), oligonucleotides were designed and a fragment of 1.5 kb of genomic sequence was amplified from genomic DNA prepared from an Arabidopsis cell-suspension culture. This DNA fragment was used to screen a genomic lambda bank prepared with DNA from the same cells, permitting the identification of two lambda clones with overlapping inserts spanning approximately 22 kb of genomic DNA and including the Arabidopsis RAD50 homologue locus. Sequencing of 10 kb from the inserts of these clones showed that they spanned the entire AtRAD50 gene (data not shown). Our genomic DNA sequence has recently been confirmed by (and is identical to that of) the Arabidopsis genome sequencing project (Lin et al., 1999).

The cDNA encoding the Arabidopsis RAD50 homologue was isolated by RACE-PCR with the Marathon Kit (Clontech Inc., Palo Alto, CA, USA). Poly(A)+ RNA was prepared from total RNA isolated from the A. thaliana cell

suspension; double-stranded cDNA was synthesized and adapters were ligated to the two ends. This library of adapter-ligated cDNA was PCR amplified using a RAD50 specific primer and primers against the adapter sequences. A PCR fragment of 4.3 kb was isolated on agarose gel and cloned into pGEM-Teasy (Promega Inc., Madison, WI, USA). Sequencing of the 4305 bp insert clone confirmed that it includes the entire AtRAD50 cDNA, and that this cDNA corresponds to the genomic clones (above). The gene covers 8486 bp and contains 27 exons; all 5' donor sites contain the AG/GT conserved junction; and the conserved AG is present in all the 3' acceptor sites. We have submitted this mRNA sequence to GenBank (Gallego et al., 1999). The computer prediction of the mRNA sequence of this gene from the genome sequencing project (Lin etal., 1999) misplaces a number of the intron-exon borders, and thus predicts a protein with 53 inserted and 32 deleted amino acids relative to that predicted from our cDNA sequence.

Amino acid comparison of Rad50 homologues.

The cloned cDNA from Arabidopsis presents a putative methionine initiation codon at nucleotide residue 146 in the first exon. No other open reading frames were detected. The termination codon of the open reading frame is located at nucleotides in position 4097, giving a predicted protein of 1316 amino acids. A 3'-untranslated region covers 181 nt before the short poly(A) tail present in the cDNA clone. The length of the protein is strongly conserved in all organisms in which it has been studied: the human, mouse and yeast Rad50 proteins have 1312 amino acids, while the C. slegans Rad50 has 1298 and the Arabidopsis RAD50 cDNA reported here predicts a protein of 1316 amino acids. Amino acid sequence comparison with the known Rad50 proteins shows a high conservation at the N- and C-terminal regions. In particular, the aminoterminal 190 amino acid region of the Arabidopsis protein has 56 and 52% sequence identity with the yeast and human Rad50 proteins, respectively. The carboxy-terminal 207 amino acids show >62 and 52% identity with the corresponding region present in the yeast and human Rad50 proteins, respectively (Figure 1a). The overall amino acid identity of the predicted Arabidopsis protein is of 29.8% with the human and 27.3% with the yeast proteins, respectively. The Arabidopsis Rad50 protein predicted from the cDNA sequence contains 18.2% of acidic amino acids and 16.9% of basic residues. The protein is predominantly hydrophilic, with 30.3% of hydrophobic residues. It presents three glycosylation motifs in positions 398-399, 419-422 and 620-623. A type-A ATP-binding side is present in the amino-terminal region at amino acid positions 34-41. A Walker B motif is present at the carboxy-terminal region between amino acids 1235 and 1242. Checking of

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the predicted AtRad50 amino acid sequence for plant localization signals with the PSORT program (http://psort.nibb.ac.jp/), yields a putative nuclear localization signal with certainty of 0.350 (success). No chloroplast or mitochondrial localization sequences were identified in the sequence by PSORT. Figure 1(b) shows the output from the coils program (http://ulrec3.unil.ch/software/ coils_FORM.html; Lupas et al., 1991). It can be seen that the predicted AtRad50 protein conserves the central coiled-coiled conformation domain seen in the yeast and human proteins (Dolganov et al., 1996).

Chromosomal localization of Arabidopsis RAD50 locus

Southern analysis of DNA prepared from Arabidopsis suspension cells show a unique band after digestion with six different restriction enzymes (Figure 2a), suggesting that the Arabidopsis RAD50 gene is present as a singlecopy gene within the A. thaliana genome.

Using PCR amplification of DNA prepared from the Arabidopsis YAC bank (Creusot et al., 1995), we have mapped the RAD50 gene on chromosome II (YAC CIC 11E1L) near the marker TEn5 (data not shown). Recently, genomic sequencing results of Arabidopsis chromosome II BAC F22D22 has confirmed this mapping (Lin et al., 1999).

Expression of the RAD50 gene in Arabidopsis

Northern analysis of RNAs prepared from Arabidopsis suspension cells as well as different plant tissues shows a single mRNA species, the length of which corresponds to that of the cDNA (Figure 2b). This has been confirmed by Northern analysis of poly(A)+ RNA from suspension cells and flower buds, as well as by RT-PCR analysis (data not shown).

Studies of human and mouse RAD50 gene expression have shown the presence of alternative spliced transcripts (Kim et al., 1996; Kim et al., 1999). We were unable to detect alternative transcripts in A. thaliana using both Northern and PCR analysis. The Arabidopsis RAD50 mRNA is expressed in all cell tissues analysed- however, stronger levels were found in fast growing cells such as cellsuspension culture, young primary roots and flowering structures. Tissue from flower buds and mature flowers contains a relatively high proportion of cells undergoing division, processes that appear to be correlated with a high level of expression of RAD50. Maximum expression of human and mouse RAD50 genes has been detected in the testis (Dolganov et al., 1996; Kim et al., 1996). Thus, although the transcript is enriched in meiotic tissues, expression of the AtRAD50 gene is not specific to meiotic cells. This pattern of expression is reminiscent of that previously reported for the Arabidopsis RAD51 homologue

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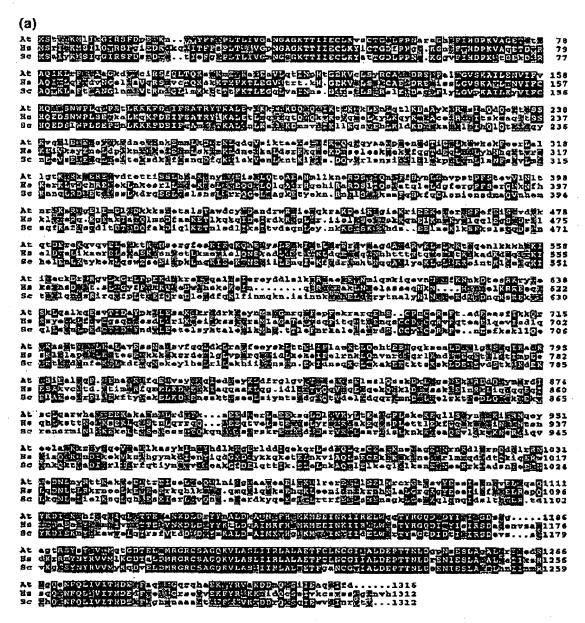


Figure 1. Comparison of AtRad50 and other Rad50 proteins.

(a) Alignment of Rad50 homologues from Arabidopsis (At), man (Hs) and S. cerevisiae (Sc). Identical residues between two or more sequences are printed as white, upper-case letters outlined in black; conservative substitutions as black, lower-case letters outlined in grey; non-conserved residues are in lower case on white. Numbers to the right of each line indicate the position in the armino acid sequence of the last residue on that line. The sequences were aligned with the clustalx program using the elections weighting matrix (Thompson et al., 1994).

(b) Comparison of predicted collect-call structure of the Arabidopsis (1316aa), human (1312aa) and S. cerevisiae (1312aa) Rad50 proteins. Output of the

(b) Comparison of prodicted collect-coil structure of the Arabidopsis (1315ea), human (1312ea) and S. cerevisiae (1312ea) Red50 proteins. Output of the COILS program (http://ulrec3.unil.ch/software/coils_FORM.html) with window width = 28; Lupas et al., 1991).

(Doutriaux et al., 1998) and those of the UVH1/AtRAD1 (Gallego etal., 2000; Liu etal., 2000) and UVR2 (Ahmad et al., 1997) genes of Arabidopsis.

As reported for the human Rad50 protein following treatment with ionizing radiation (Dolganov et al., 1996), the steady-state level of AtRAD50 mRNA does not change in cultured cells in response to exposure to the radiomimetic compound MMS (data not shown). This contrasts with the situation reported for mouse RAD50 gene expression, which does increase after treatment of NIH-3T3 cells with MMS (Kim et al., 1996).

Identification and characterization of a rad50 mutant of Arabidopsis

Based on our AtRAD50 sequence, we designed oligonucleotides and screened the Versailles Arabidopsis T-DNA insertion mutant collection (Bechtold et al., 1993; Bouchez et al., 1993) using PCR as described by Gaelen et al. (2000). From this screen we identified a single plant containing a T-DNA insertion in the RAD50 locus. The mutant plant is fully sterile, producing numerous flowers and small, empty siliques (Figure 3).

The mutant phenotype and the kanamycin resistance marker of the inserted T-DNA co-segregate, with selfed heterozygotes producing progeny with the 3:1 ratio expected for a single-locus insertion (128 non-mutant : 38 mutant; χ^2 , 1 df = 0.39). Southern analysis has confirmed that the T-DNA insertion is present at a single locus (not shown), and PCR results indicate the single-copy nature of the insertion (see the size of the amplified fragment present in the homozygous plants, Figure 4). Molecular analysis shows that the 38 plants presenting the sterility

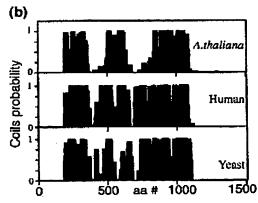


Figure 1b. Logand on facing page.

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phenotype are, as expected, homozygous for the T-DNA insertion. Thus there is clear linkage between the T-DNA insertion and the sterility phenotype of the mutant rad50 allele, which is recessive. Furthermore, we have also observed a sterility phenotype in plants expressing AtRAD50 antisense RNA (not shown). These results suggest a role for the Rad50 protein in meiosis in plants which is consistent with that known in yeast. Due to the cell-lethal phenotype of the rad50 null mutant in mouse, this constitutes the first implication of RAD50 in meiosis in a metazoan organism.

Mutant plants which have been germinated on agar medium before being transferred to soil are reduced in size in comparison to their non-mutant siblings; however this

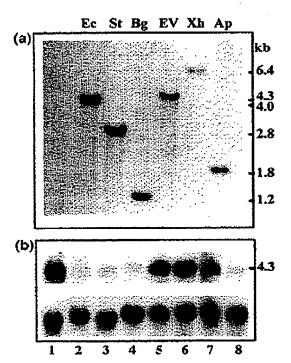
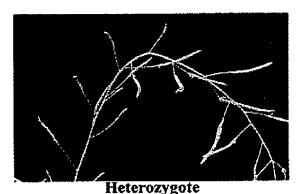


Figure 2. Southern and Northern analysis of AtRAD50. (a) Southern analysis of DNA from Arabidopsis cell culture probed with the ArRAD50 genomic probe. From left to right: EcoR1+, Sty1+, Byth-, EcoRV-, Xhol- and Apal-digested DNA. Sizes of the detected fragments are shown to the left.

(b) Northern analysis of retal Arabidopsis RNA from different tissues, probed with the 3' half of the AtRAD50 cDNA (upper) and the 185 rDNA probe as a loading control (lower). Lanes: 1, suspension cells; 2, young roactto leaves grown for 3 weeks; 3, leaves from stems of moture flowering plants; 4, rosotto looves from non-llowering plants grown for 6 weeks with short daylength (8 h light period); 5, mature flowers; 6, flower buds; 7, roots; 8, stems.

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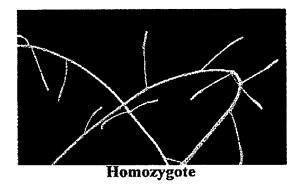


Figure 3. The rad50 mutant is storile.
Phatographs of 7-week-old mutant (rad50/rad50) and heterotygote(rad50/RAD50) plants. The mutant plants produce many flowers and small, empty siliques.

c d

b

Figure 6. MMS sensitivity of rad50 mutant cells.
Mutant (rad50/rad50 upper on each plate) and heterozygote (RAD50/rad50 lower on each plate) suspension cell lines grown in Potri dishos on agar medium containing different concentrations of MMS: (a) no MMS: (b) 0.0033%; (c) 0.0067%; (d) 0.01%; (e) 0.0133%; (f) 0.0167% v/v MMS.

phenotype is not seen if seeds are germinated directly in soil in the greenhouse. This growth difference is seen whether or not antibiotic (kanamycin) is included in the agar medium, and is thus not an artefact caused by the antibiotic. Further studies will be needed to determine the exact origin of this facultative growth defect.

The mutated rad50 gene and its inserted T-DNA were PCR-amplified, cloned, and the plant DNA/T-DNA junctions sequenced to determine the exact position and structure of the insertion (Figure 5). The T-DNA is Inserted into the coding sequence of the gene and causes disruption of the RAD50 ORF in the 21st exon, after the 1050th codon (of 1316). Thus any mutant protein produced would lack the 266 C-terminal amino acids of the wild-type protein.

The inserted T-DNA has a small, 141 nucleotide, inverted duplication of the LB sequence upstream of the RB, and has lost the first 10 nucleotides following the RB nick site. The inverted orientation of this LB DNA implies the participation of a replicated T-DNA molecule in the origin of this sequence, and that it is not simply the result of

ligation-derived concatamers of the single-stranded transforming T-DNA molecules. The beginning of the insertion (inverted LB sequence) has a 12 bp (with three mismatches) homology to the RAD50 sequence at that position. The LB junction is much simpler, with the T-DNA sequence ending 16 nt downstream of the LB nick site, followed by the insertion of two As prior to resumption of the RAD50 gene sequence. The Insertion caused an 18 bp deletion (not including the 12 bp homology, above) of RAD50 sequence. Neither of the two junctions occurs exactly at the nick site of the corresponding border sequence. We have previously seen similar structures at T-DNA insertion sites (unpublished results), and the mechanism of T-DNA integration has been reviewed by Tinland (1996). The systematic sequencing of the DNA flanking T-DNA insertions of the Versailles mutant collection is currently under way, and understanding of these events will greatly benefit from this rich source of data.

The Arabidopsis rad50 mutant is hypersensitive to MMS

In yeast cells, mutation of the RAD50 gene causes hypersensitivity to X-ray irradiation and to particular radiomimetic DNA-damaging chemicals such as MMS. However, due to the cell-lethality of rad50 mutants in

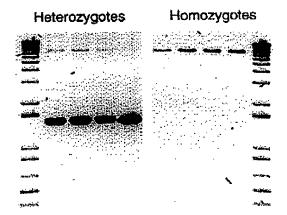


Figure 4. The rad50 mutant facus contains a single T-DNA insertion. PCR of individual plants segregating RAD50. Total DNA was amplified with a pair of primers in the RADSO sequence, spanning the T-DNA insertion site (5'-GAGCTGTGAAGCTAGAAAGAATGAACTTGCAGGTG: 5'-CCCATCCAGGTTTGTAGTTG). Homozygote plants show only the 8 kb PCR product expected for a single-copy T-DNA insertion, while heterozygotes also show the expected 1.5 kb band for the wild-type RAD50 gene.

mouse, a clear demonstration of this radiosensitivity has not previously been possible in a metazoan organism (see Luo et al., 1999 for work on gamma-irradiated early mouse blastocyst explants).

We and colleagues (J. Paszkowski, personal communication) have observed that the lethal effects of MMS on Arabidopsis seedlings are dependent on the size of the plants tested. Non-ambiguous determination of the MMS concentration needed to kill a given plant line (or other multicellular organism) is thus complicated by questions of the size of plants used. In order to avoid this problem, we therefore tested MMS sensitivity on cell lines derived from these plants. In this way, very small clumps (microcalluses) of growing cells are exposed to MMS, and a much clearer determination of dose-dependent MMS sensitivity is possible. Callus was induced on young leaf tissue, and suspension cultures initiated using standard protocols (see Experimental procedures). Suspension cells in liquid culture were pipetted onto solid growth media with or without different concentrations of MMS, and their growth scored visually 2-3 weeks later (Figure 6). In the absence of MMS the rad50 mutant and heterozygote cell lines had both grown to similar extents, whilst the rad50 mutant cells showed a clear hypersensitivity to MMS relative to the heterozygote cell line. We have also observed MMS sensitivity in mutant cell lines generated via the antisense approach (data not shown). These data suggest that the Rad50 protein plays a role in DSB repair in plant cells, as is the case in yeast.

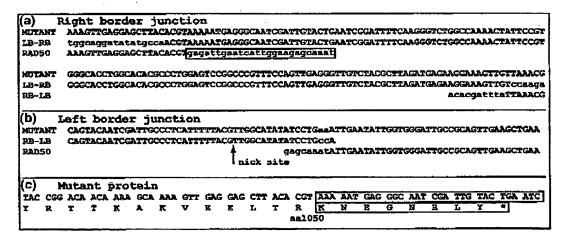


Figure S. Sequences of junctions of the inserted T-DNA RAD50 junctions (a,b) and the predicted effect on the Rad50 protein (c). The mutant rad50 genomic DNA sequence is aligned with the sequences of the ends of the T-DNA used to generate the Anabidopsis mutents, and with the RAD50 genomic sequence. RAD50 sequence deleted at the point of the insertion is boxed (a) and the RB-LB sequence shown begins immediately after the RB nick (b). In frame coding DNA (upper) and protein flower) sequence added to the mutant rad50 open reading frame by the insertion is shown boxed in (c).

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The Arabidopsis rad50 mutant is viable

To our knowledge, the only other report of a rad50 mutant in a metazoan organism describes a 'cell-lethal' phenotype in a mouse rad50 mutant (Luo et al., 1999). These authors were unable to isolate homozygous rad50 mutants of mouse ES cells in culture, and ascribed the early embryonic lethality of homozygous rad50/rad50 mice to a gradual cession of proliferation of the embryonic cells. A differential radiosensitivity was also seen in mutant blastocyst explants, and the authors tentatively concluded that the cell lethality is caused by the accumulation of unrepaired and/or misrepaired DNA DSBs, indicating involvement of mouse Rad50 protein in DSB repair/ signalling.

We show here that the Arabidopsis rad50° mutant is hypersensitive to the radiomimetic agent MMS, and thus presumably defective in DSB repair; however we have no evidence for a strong effect on the proliferative capacity of rad50° Arabidopsis cells. Under certain growth conditions the mutant plant is stunted, but the facultative nature of this phenotype clearly indicates the lack of a basic problem at the cell division level. As the mutant plant is fully sterile, effects on cellular proliferative capacity can only be studied in the context of a single plant generation. Thus we have derived mitotic cell cultures from the mutant plants in order to be able to study long-term effects. These cells grow, and we are undertaking further analysis on the involvement of the RAD50 function in DNA repair/recombination/cell proliferation in plant cells.

As the rad50 mutation in our mutant is caused by a DNA insertion into the coding sequence of the AtRAD50 gene, we cannot exclude that a truncated protein is produced in the mutant cells, which would thus not be true 'null' mutants. Preliminary data using antibodies to a C-terminal fragment of the AtRad50 protein shows the absence of the AtRad50 protein band on Western blots of protein from the mutant cells (S. Daoudal and C.I.W., unpublished results); however these antibodies may not recognize a C-terminustruncated form of the protein. It is thus formally possible that this putative truncated protein is sufficient to fulfil the function of the Rad50 protein in cell viability. However, given the clear phenotypes of the mutant for meiosis and MMS sensitivity, we think it extremely unlikely that the viability of the rad50⁻ mutant cells and plants can be explained in this way. We note that the question of whether a given mutation is null may be posed for any insertional mutation that does not involve full deletion of the open reading frame of the gene in question (i.e. including all insertional mutations). Whether or not our mutant is 'null' does not alter our conclusions on the role of the AtRad50 protein in Arabidopsis and the phenotypes of the T-DNA insertion mutant described here. Full resolution of this question must await the identification of a full Atrad50 deletion mutant.

In conclusion, the plant Rad50 function appears to be analogous to that of yeast. Given the difficulties encountered in isolating and studying the mouse rad50 mutant, we believe that further study of rad50 mutant plant cells will permit us to clarify the role of this polyvalent protein and its multiple roles in the maintenance of genome integrity in both plants and animals.

Experimental procedures

Growth of cell suspensions and plants

The Arabidopsis thaliana cell suspension (TB7) was established by Axelos et al. (1992). Cells were grown in Gamborg's B-5 medium (Sigma #G5893, St Quentin Fallavier, France) supplemented with 30 g l $^{-1}$ sucrose and 200 mg Γ^{-1} naphthalene acetic acid (NAA) on a rotating platform (120 rpm) at 22°C with 16 h light/8 h dark. Cells in liquid culture were subsultured at weekly intervals. Bacto-agar (Difco, Detroit, MI, USA) was included in the cell-suspension medium at 0.8% w/v for solid media. Arabidopsis thaliana (Columbia) seeds were sown directly into damp compost and germinated in a greenhouse under white light (16 h light/8 h dark). For the isolation of root tissue, plants were grown aeroponically under the same conditions.

Callus cultures were derived from homozygous rad50/rad50 and heterozygous rad60/RAD60 plants using standard techniques as follows (J. Lucht and B. Hohn, personal communication). Leaves of young germinating plants were surface-sterilized in 0.5% sodium hypochlorite, 0.05% Tween-80 for 10 min at AT and rinsed several times with sterile water. The leaves were then cut up with a sterile scalpel and placed on callus induction medium (CIM) agar for 1 week (22°C, 16 h light). The leaf/callus was then transferred to shoot induction medium (SIM) agar medium, and after 2-3 weeks successfully growing green callus was transferred to fresh SIM medium (either solid or liquid) and then maintained on this modium by regular subculture. CIM and SIM media were prepared as for the Gamborg's B-5 medium (above), except that the hormones (Sigma) differ: CIM, 1 mg F1 2,4-dichlorophenoxyacetic acid, 0.2 mg F¹ kinstin; SIM, 0.1 mg F¹ NAA, 1 mg F¹ 6benzylaminopurine.

Isolation of the AtRAD50 cDNA and genomic DNA clones

Using poly(A)+ RNA prepared from the cell-suspension culture of Arabidopsis, the complete AtRAD50 cDNA was isolated using the marathon RACE PCR kit following the manufacturer's instructions (Clontech).

The genomic DNA clone was isolated from a genomic DNA library constructed with Sau3A1 partially digested DNA from the cell suspension cloned into lambda FIXII (Stratagene Inc., La Jolla, CA, USA) following the manufacturer's protocol.

Southern analysis

Genomic DNA was isolated from cell-suspension cultures following the method of Dallaporta et al. (1983). DNA (3 µg) was digested with 50 units of the relevant restriction enzyme in a volume of 100 µl for 16 h at the recommended temperature.

Digested samples were phenol/chloroform extracted, ethanol precipitated, resuspended in TE and electrophoresed in 0.8% agarose/TAE gels. The gels were capillary blotted to Hybond N+ (Amersham, Orsay, France) positively charged nylon membrane, and hybridized at 62°C to radioactively labelled DNA probes according to Church and Gilbert (1984). Filters were then washed (0.1 × SSC, 0.1% SDS, 62°C) and autoradiographed. Probes were labelled with a-32P dCTP using the Prime-It II kit (Stratagene) according to the manufacturer's instructions.

RNA isolation and Northern analysis

Frozen plant or cell-culture tissue was homogenized in liquid N2 with a mortar and pestle. Total RNA was then prepared using the Trizol reagent following the manufacturer's instructions (Gibco-BRL, Cergy Pontoise, France). Poly(A)+ RNA was purified from total RNA using the mRNA Direct kit (Dynal Inc., Compiègne, France). For Northern analysis, 30 µg RNA per lane was fractionated on 0.8% agarose/formaldehyde gels, which were blotted, hybridized to radioactively labelled probes, and autoradiographed as for the Southern blots (above).

Callus MMS sensitivity tests

0.5 ml of suspension cells were pipetted onto the surface of agar plates containing solid SIM medium and different concentrations of MMS (Sigma #M4015). The plates were then incubated as described above, and resistance or sensitivity was scored visually 2-3 weeks later. MMS-containing plates were prepared immediately before use. All MMS-contaminated material was quenched after use by soaking in 10% w/v sodium thiosulphate.

Acknowledgements

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APPENDIX D

APPENDIX D - motifs from Example 4, Pfam added to Appendix A (2/14/02) Docket 1116B Serial Numb r 09/538,396

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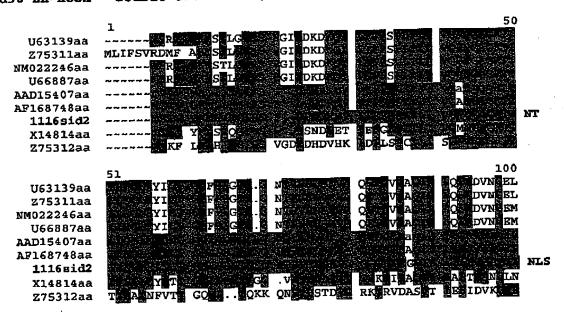
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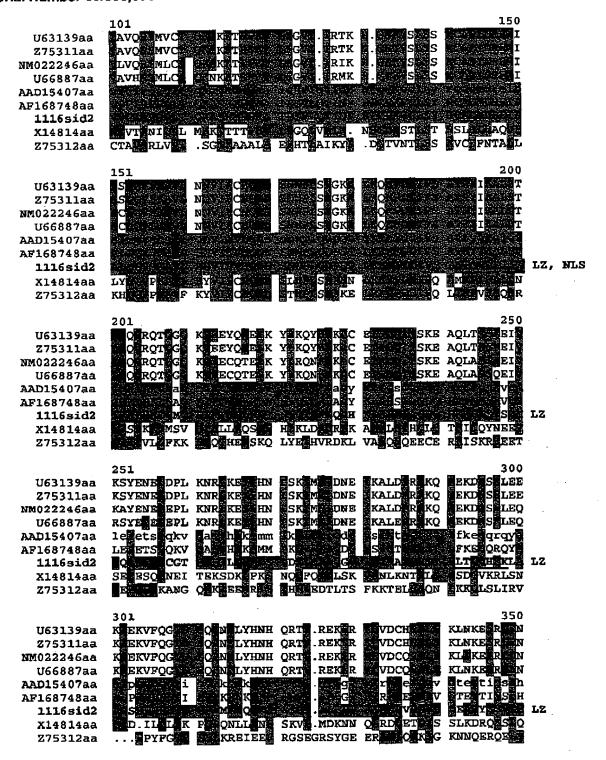
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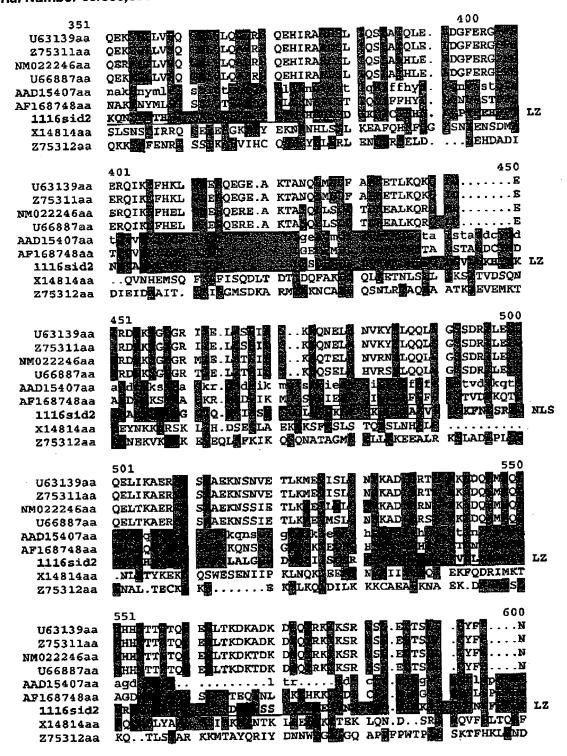
MOTIFS: Identified in Example 4 and Pfam analysis Leucine Zipper - Underlined, LZ noted in left margin Nucleotide binding motif - Italicized & underlined, NT in left margin Nuclear localization signal - boxed, NLS in left margin Rad50 Zn-Hook - Double-underlined, ZN noted in left margin



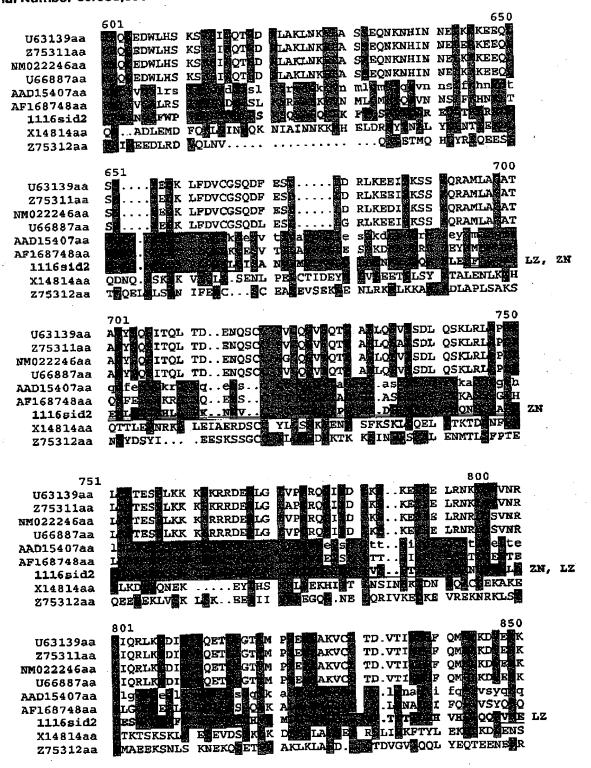
APPENDIX D - motifs from Example 4, Pfam added to Appendix A (2/14/02) Docket 1116E Serial Number 09/538,396



APPENDIX D - motifs from Example 4. Pfam added to Appendix A (2/14/02)
Docket 1116E
S rial Number 09/538,396



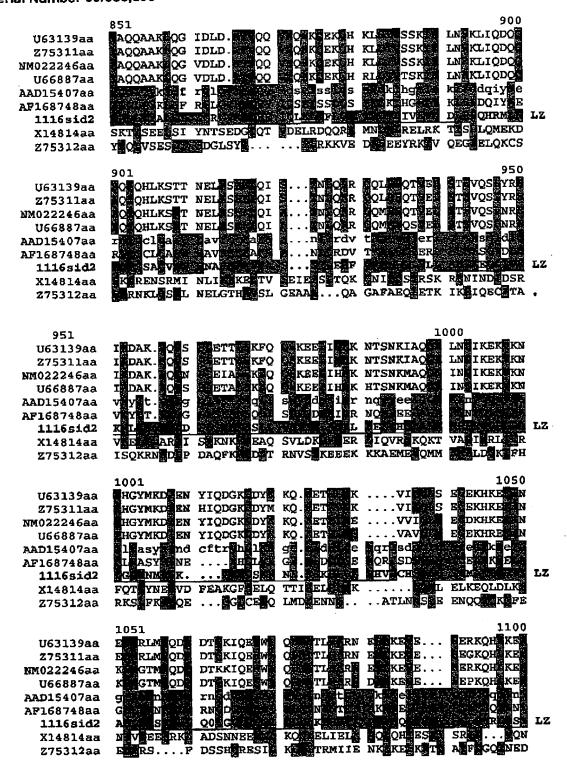
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Docket 1116E
Serial Number 09/538,396



APPENDIX D - motifs from Example 4, Pfam added to Appendix A (2/14/02)

Docket 1116E

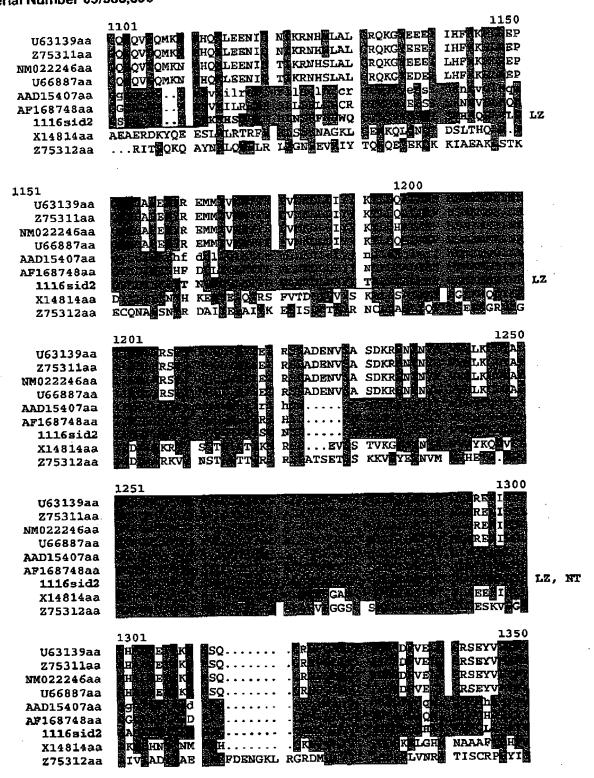
Serial Number 09/538,396



APPENDIX D - motifs from Example 4, Pfam added to Appendix A (2/14/02)

Docket 1116E

Serial Number 09/538,396



APPENDIX D - motifs from Example 4, Pfam added to Appendix A (2/14/02) Docket 1116E Serial Number 09/538,396

1351			80
U63139aa	KENIDEC	ENVKCS	
Z75311aa	KKNIDC	EVKCS	
NM022246aa	KINIDC		GSYVH
U66887aa	KINMDIC	E VKCS SSL	GSYVH~~~
AAD15407aa	m~~~	~~~~~~~	~~~~~~~
AF168748aa		radig angawi –	~~~~~~
1116sid2	N	~	~
X14814aa	K R K	O WV NRV	TY~~
Z75312aa	CLG HGI	FESKRYPDGT	VKRVNTKRRF

Page 1 of 5



Projects Analyses Admin Alert SRS Tutorial

Analysis Browser: Level Up

Report for

1116E.rad50.sid2 (Protein)

Update

Description

1116E_rad50_sid2

Edit

Function

DNA repair protein RAD50 (153 kDa protein).

Direct assignment of functionality by homology to

swiss|P12753|RA50_YEAST

in region 1 to 1314 for overall length of 350 (99% of query, 375% of hit, see the

alignment).

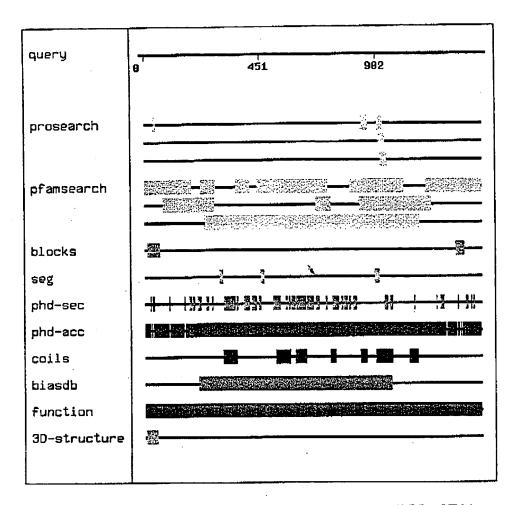
Functional class

Transcription

Extracted keywords

ATP-binding, Coiled coil

Features Summary bioSCOUT 1.5.3 - feature report for 1116E.rad50.sid2



Homologies			All BLAST hits
	Protein	30 clear homologs	All protein BLAST hits
	ESTs	170 homologs	All EST BLAST hits
	Patents	53 homologs	All patent hits
General	Gene name		
	Molecular weight	152.50 kD	
	Sequence length	1316	
	Isoelectric point	6.04	
	Predicted cellular localisation (PHD and PreLoc)	nuclear (94.7 %)	

3D Structure

3D structure inferred by unlikely homology from residues 6 to 48 in 1E69-A

bioSCOUT 1.5.3 - feature report for 1116E.rad50.sid2

Page 3 of 5

	·	<u>alignment</u>
	pdb <u> 1E69 1E69-A</u>	<u>structure</u>
Phylogeny	Distribution	34 species extracted from 175 Species homologous sequences.
	Taxa	Chordata, Eukaryotae, Fungi, Planta
	Model organisms	Arabidopsis thallana, Caenorhabditis elegans, Drosophila melanogaster, Homo sapiens, Mus musculus, Saccharomyces cerevisiae
Features	Coiled coil region	from 311 to 336, from 341 to 361, from 517 to 538, from 541 to 569, from 592 to 633, from 727 to 747, from 844 to 867, from 903 to 928, from 930 to 969, from 1036 to 1066 detected by [Coils]
	Low complexity region	from 296 to 307, from 456 to 469, from 902 to 915 detected by [seg]
	E-rich region	from 212 to 964 detected by [biasdb]
	No significant hits detected by	[Phd-tm]
Patterns	SMC family, C- terminal domain - region	from residue 1100 to 1314. Source: [pfamsearch]. Quality: (E=0.52)
•	ABC transporter - region	from residue 1141 to 1296. Source: [pfamsearch]. Quality: (E=0.19)
	Intermediate filament protein -	from residue 840 to 1118. Source: [pfamsearch]

from residue 237 to 1075. Source: [pfamsearch] Myosin tail - region from residue 804 to 1015. Source: [pfamsearch]

Uncharacterized ACR, COG1579 region

region

KE2 family protein -

region

Protein of unknown

Rad50 zinc hook motif region

from residue 673 to 726. Source: [pfamsearch].

from residue 891 to 983. Source: [pfamsearch]

Quality: (E=9.9e-06)

from residue 441 to 717. Source: [pfamsearch]

bioSCOUT 1.5.3 - feature report for 1116E.rad50.sid2

function, DUF259 region

Late embryogenesis abundant (LEA)

from residue 548 to 625. Source: [pfamsearch]

group - region Rad50 zine hook

from residue 360 to 413. Source: [pfamsearch]

motif region Heat shock protein

from residue 219 to 275. Source: [pfamsearch]

9 / 12 - region Poly(A) polymerase

from residue 75 to 273. Source: [pfamsearch]

central domain region Sigma-70, non-

from residue 112 to 260. Source: [pfamsearch]

essential region region

from residue 2 to 182. Source: [pfamsearch]. Quality:

RecF/RecN/SMC N terminal domain region

(E=0.48)

LEUCINE_ZIPPER region

from residue 922 to 944. Source: [prosite]

from 915 to 937. Source: [prosite]

from 908 to 930. Source: [prosite] from 849 to 871. Source: [prosite]

ATP_GTP_A region

from residue 34 to 42. Source: [prosite]

No significant hits found in

[blocks database]

Comment

No comment section.

Edit.

Completed Tasks

Start Time

Comment

Output

Interactive

04.04.2003, dressym

User

bioSCOUT default details...

16:44:19

Permissions

Edit

Alert Jobs

New:



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Alignment: 1116E.rad50.sid2 - pdb|1E69|1E69-A

BLASTP - alignment of 1116E.rad50.sid2 against pdb|1E69|1E69-A

chromosome segregation smc proteinfragment: smc fusion of the n- and c-terminal globular domains residues 1-152 and 1023-1164;

- This hit is scoring at: 0.56 (expectation value)
- Alignment length (overlap): 43
- Identities: 44 %
- Scoring matrix: BLOSUM62 (used to infer consensus pattern)
- Database searched : nrdb

Q:	6 KML1KGIRSFDPDNKNVITFFKPLTLIVGPNGAGKTTIIECLK				
_		K:.:KG.:SF:: I F . :T.IVGPNG:GK:.II:.:K			
H:	5	KLYLKGFKSFGRPSLIGFSDRVTAIVGPNGSGKSNIIDAIK	45		

Legend of Alignment

- : positive score
- . score between -2 and 0

Entry Page

```
1£69
                                                      09-AUG-00
          CHROMOSOME SEGREGATION
HEADER
TITLE SMC HEAD DOMAIN FROM THERMOTOGA MARITIMA
          MOL ID: 1;
COMPND
COMPND 2 MOLECULE: CHROMOSOME SEGREGATION SMC PROTEIN;
COMPND 3 CHAIN: A, B, C, D, E, F;
COMPND 4 FRAGMENT: SMC FUSION OF THE N- AND C-TERMINAL GLOBULAR
         2 MOTECULE: CHROMOSOME SEGREGATION SMC PROTEIN;
COMPND 5 DOMAINS RESIDUES 1-152 and 1023-1164;
COMPND 6 ENGINEERED: YES SOURCE MOL ID: 1;
SOURCE 2 ORGANISM SCIENTIFIC: THERMOTOGA MARITIMA;
SOURCE 3 ATCC: DSM3109;
SOURCE 4 CELLULAR_LOCATION: CYTOSOL;
SOURCE 5 EXPRESSION_SYSTEM: ESCHERICHIA COLI;
SOURCE 6 EXPRESSION_SYSTEM_STRAIN: C41(DE3);
        7 EXPRESSION_SYSTEM_CELLULAR_LOCATION: CYTOSOL;
SOURCE
SOURCE 8 EXPRESSION_SYSTEM_VECTOR_TYPE: PLASMID;
SOURCE 9 EXPRESSION_SYSTEM_PLASMID: PHIS17
        SMC, STRUCTURAL MAINTENANCE OF CHROMOSOMES, COILED COIL
KEYWDS
         X-RAY DIFFRACTION
EXPDTA
           J.LOWE, S.C.CORDELL, F. VAN DEN ENT
AUTHOR
REVDAT 2 27-MAR-01 1E69 1
REVDAT 1 09-AUG-00 1E69
                                 0
            AUTH J.LOWE, S.C.CORDELL, F. VAN DEN ENT
JRNL
             TITL CRYSTAL STRUCTURE OF THE SMC HEAD DOMAIN: AN ABC
JRNL
             TITL 2 ATPASE WITH 900 RESIDUES ANTIPARALLEL COILED-COIL
JRNL
JRNL
             TITL 3 INSERTED
JRNL
                                                      v. 306
             REF
                    J.MOL.BIOL.
             REFN ASTM JMOBAK UK ISSN 0022-2836
JRNL
```



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Features

Summary

Searched query 1116E.rad50.sid2 against PFAM database.

Hit	Score	Expect	Description	Q from	Q to	Method
∏ <u>pfam hmm Rad50_zn_hook</u> . <u>align</u> ment	32.5	9.9e- 06	Rad50 zinc hook motif	673	726	HMMPFAM
pfam hmm Rad50 zn hook . alignment	4.2	1.8	Rad50 zinc hook motif	360	413	НММРГАМ
pfam hmm HSP9_HSP12.alignment	-11.4	4.1	Heat shock protein 9 / 12 -	219	275	HMMPFAM
F pfam hmm LEA_1. alignment	-14.3	5.6	Late embryogenesis abundant (LEA) group -	548	625	HMMPFAM
□ pfam hmm KE2. alignment	-19.0	5.6	KE2 family protein -	891	983	HMMPFAM
pfam hmm PAP_central.alignment	-45.9	7.6	Poly(A) polymerase central domain -	75	273	HMMPFAM
pfam hmm ABC_tran.alignment	-47.7	0.19	ABC transporter -	1141	1296	HMMPFAM
pfam hmm SMC_N . alignment	-69.5	0.48	RecF/RecN/SMC N terminal domain -	2	182	HMMPFAM
T pfam hmm DUF164. alignment	-90.1	2.8	Uncharacterized ACR, COG1579 -	804	1015	HMMPFAM
Γ <u>pfam hmm SMC_C</u> . alignment	- 116.7	0.52	SMC family, C- terminal domain -	1100	1314	HMMPFAM
□ pfam hmm sigma70_ner.alignment	- 122.2	4.9	Sigma-70, non- essential region -	112	260	HMMPFAM

pfam hmm DUF259. alignment	- 137.1	9.3	Protein of unknown function, DUF259 -	441	717	HMMPFAM
☐ pfam hmm filament . alignment	203.9	5.4	Intermediate filament protein -	840	1118	HMMPFAM
☐ pfam hmm Myosin_tail alignment New Task Rename S	555.1 equences	9.8	Myosin tail -	237	1075	HMMPFAM



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				.:	Features

Alignment: 1116E.rad50.sid2 - pfam|hmm|Rad50_zn_hook

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|Rad50_zn_hook

Rad50 zinc hook motif

• This hit is scoring at: 4.2

Scoring matrix: BLOSUM62 (used to infer consensus pattern)

Q:	360 AHLTMKHERDSDIKNICTKHNLGPVPEHPF-TNDVAMNLTNRAKARLSSLENDLL	413
	. L:	
н:	1 galesekaelkkaieeleeesscCPvCgReLgteeekkelikeykseldrlpeelk	57

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|Rad50_zn_hook

Rad50 zinc hook motif

• This hit is scoring at: 32.5

Scoring matrix: BLOSUM62 (used to infer consensus pattern)

Q:	673 NFANGMREMLAPFEHLARKNHVCPCCERAF-TPDEEDEFVKKQRMQNSSTAERSK	726
_	:. L: CP.C R T.:E:.E.:K:.:.: K	
H:	1 galesekaelkkaieeleeesscCPvCgReLgteeekkelikeykseldrlpeelk	57

Legend of Alignment

positive score

score between -2 and 0



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Alignment: 1116E.rad50.sid2 - pfam|hmm|HSP9_HSP12

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|HSP9_HSP12

Heat shock protein 9 / 12 -

• This hit is scoring at: -11.4

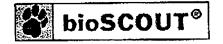
• Scoring matrix: BLOSUM62 (used to infer consensus pattern)

Q:	219 —-DQAHKLRENIAQDQEKSDASKSQMEQLKEKICGTEREILQMETSLDELRRLQGQIDI			
_	DA.K . A:::D:SKSEQ:KEK: :: TS D::Q . D			
H:	<pre>1 MSD1aRKdFgeKakEklTPDSsKSTaEqvKEkvTDklDkvAgkvtsdddKStvQkAhDk</pre>	59		

Legend of Alignment

: positive score

. score between -2 and 0



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Features

Alignment: 1116E.rad50.sid2 - pfam|hmm|LEA_1

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|LEA_1

Late embryogenesis abundant (LEA) group -

- This hit is scoring at: -14.3
- Scoring matrix: BLOSUM62 (used to infer consensus pattern)

Q: 548 LESSKDKLNEIVNEHKDKIKKVLrgrnpFEKDMKKEINQAFWPVBKEYNELRSKSQEAEQ
::S:K:K:::: K:K: : D K.E .A .:KE. . R.K::EA:
H: 1 MqSaKEKisnmAstaKekmditK....AkadEKaEkatARtkeEkelAhqrkkAKeAqA

ELKFTQSKVTDAREQLTK 625 E:..::K...A.E::. eMdlheaKAehaaekesa 73

Legend of Alignment

- : positive score
- . score between -2 and 0



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			Features

Alignment: 1116E.rad50.sid2 - pfam|hmm|KE2

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|KE2

KE2 family protein -

- This hit is scoring at: -19.0
- Scoring matrix: BLOSUM62 (used to infer consensus pattern)

Q:	891	ASSILERFOKSEEELVLLAEEKEOLIVEKKLLEESLDPLSKEKE3	LT-Q
		:L.:.Q: :::L ::K.QL : K .E L:.L.K .E.	: Q
H:	1	vqellaklqqlqqqlekvmtqkaqlerqLkEaelvleELekldeDtkVYklV	'GkvLVkvq
		EYNALKQKLDEEYHQLAERKREFQQELDALGRLNMKIKGYLDSKK ::.L:EQL.E. :::: L : L. K:: .L.S	983
		dkeeardeLeerleqleeeiktLekqeeylekeleeleeklqellqsaa	109

Legend of Alignment

- : positive score
- . score between -2 and 0

bioSCOUT 1.5.3 - Alignment: 1116E.rad50.sid2 - pfam|hmm|PAP_central

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Alignment: 1116E.rad50.sid2 - pfam|hmm|PAP_central

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|PAP_central

Poly(A) polymerase central domain -

• This hit is scoring at: -45.9

Scoring matrix: BLOSUM62 (used to infer consensus pattern)

Q: 75 XSNWPLQDPSTLKKKFDDIFSAT-RYTKALEVIKK-LÄKDQMQEIKTFRLK
:P. PSTL:KF::FS. R:...V: K::.D.::E...

H: 1 RFE.radP.lYPnavpstlvekfFlvfsqWlrhnwpnPVlLkeinsdsieernlqvrvRF

273

E.rade 205

Legend of Alignment

- : positive score
- . score between -2 and 0



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				•	Help
				14.1 2.00	Features

Alignment: 1116E.rad50.sid2 - pfam|hmm|ABC_tran

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|ABC_tran

ABC transporter -

• This hit is scoring at: -47.7

• Scoring matrix: BLOSUM62 (used to infer consensus pattern)

Legend of Alignment

- : positive score
- . score between -2 and 0



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18. A 18. 1					Help
				. *** **************************	Features

Alignment: 1116E.rad50.sid2 - pfam|hmm|SMC_N

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|SMC_N

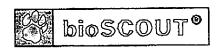
RecF/RecN/SMC N terminal domain -

- This hit is scoring at: -69.5
- Scoring matrix: BLOSUM62 (used to infer consensus pattern)

Q:	2	STVDKMLIKGIRSFDpdNKNVI-TFFKPLTLIVGPNGAGKTTIIBCLKLSCT-GEL :::::::G.:S: .K.VI .FT.IVGPNG:GK:.I:::: :::::::L
H:	1	mylkrielegFKSYagktvigpFspgFtaIvGPNGSGKSNilDAIlFvLGegrsakkL
•		PPNSRSGHTFVHDPKVAGeteTKGQIKLRFKTAAGKDVVCIRSFQLTQKASKME S A::::.F ::: IRL.:: .::. RaerlsdLIhkgsggkppanksAcVtitFdnedkeniselgqhirdgpldeenpevt
		FKAIESVLQTINPHTGEKVCLSYRCADMDREIPALMGVSKAV : I N K V A.: D E:L ItRrvyrlglgdGstSeYyiWknlrlNgkrvtklkevqelLesagIdiElatLangvsky
·		RayaWCFHWDkELiLleqIswRaeYeaLapeiETCqlFLPElLtvSfqrGWeKetdYaEv
		LENVIFVHQDE
		LarnfERDkqLgYTqGPqKADLR)RAnGlPVEDvLSRGQLKLknpyfiilOGeyltrokg
		SNWPLQDPSTLKKKFDDIF-S 182
		rhClYLvedIasmkPkeRreldeGLlellEEisGt 330

Legend of Alignment

- : positive score
- . score between -2 and 0



Alert Tutorial Admin Projects Analyses Help **Features**

Alignment: 1116E.rad50.sid2 - pfam|hmm|DUF164

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|DUF164

Uncharacterized ACR, COG1579 -

- This bit is scoring at: -90.1
- Scoring matrix: BLOSUM62 (used to infer consensus pattern)
- 804 PTDTID:HVHEIQQLVKEVEDLEYALDSSgrgVKSLEEIQLELNPLQRTRDTLIVEVDDL Q: .:.:. :: IQ:: KE E LE .:.. K.L::.: L. L::. :.L E ::L 1 mknelk.sLvkiqeidkekerLeerikei...pkeLkkakellealkkeveeleqekeel H: RDOHRMLNEDMSSAQVRWHNAreekvka-----SSILERFQKSEEELVLLAEEKEQLI .:: ...QK:::: V.L.:E EQL ::: :.L.:::...: : ..A EEK:.. koevkklekeigeieekikka. EekmdeiktqrEYkALerElqkakdkevtlrkeieqle VEKKLLEESLDPLSKEKESLLQEYNALKQKLDEEYHQLAERKREFQQELDalgRLNMKIK E K : EE .: : L .: E . : : : E . : : E . : : . L . K . . eelkkieeeieelkeeilkqEkeleeeeeevelEvrkikekvlellskre...elkektd GYLDSKKNEKLKELQGRHVL-----CH----SQLQSCMAKQ-QRIS------.. :K. :. :: CH SQ.:: :.K: . I edllsfYERiiknkknlviVPiennvCaGChiiLpsqfenkVrkePddivfCPyCSRILY 1015 AEL E yee 235

Legend of Alignment

- positive score
- score between -2 and 0



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				() ()	Features

Alignment: 1116E.rad50.sid2 - pfam|hmm|SMC_C

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|SMC_C

SMC family, C-terminal domain -

- This hit is scoring at: -116.7
- Scoring matrix: BLOSUM62 (used to infer consensus pattern)

```
1100 ISKHKQELKLS------QYKDIEKRYTNQFLQLKTTEMANKDLDRYYTALDKAL
Q:
                              :Y.::E:RY.. . :L:..E ..K.L .....LDK.
       l lekeikeLgpVlnElqvNlkAieEYaeveeRyeelvekledleeerkklleviseldkkr
H:
         {\tt MRFHSMKMEEINKIIKELWQQ-TyrgqdidyisinsdsegagtrsysyrvvmQTGDAELE}
                                                            GDAEL.
         :. .. :::INK .KE::Q: T
         leeFmeafnkInknfkevFqeLt......gGGdAeLr
         MRG---RCSAGQKVLAS-----LIIRL------ALAETFCLNCG----ILALDEPT
         :.. S:G :: A. :.L ALA .F.::
         \verb|LtDpdDPFssGieisArPPgKkwknlelLSGGEKtLtALAL1FAIhkykPsPFYvlDEvD|
         TNLDGPNAeSLAAALLRimeaRKGQENFQLIVITHDERFAhligqrQLAEKYYRVSKDEN
         . LD .M. S .A .:: R: .:N Q.IVI: .... : A:... V .:::
         AALDeaNV.sRvAnyIk....rersknaQFIVIsLRnnmm....ekADaLvGVymqdd
         QHSIIESQEI
                      1314
           S : S ::
         gvskVislkL
                       210
```

Legend of Alignment

- : positive score
- . score between -2 and 0



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					Features

Alignment: 1116E.rad50.sid2 - pfam|hmm|sigma70_ner

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|sigma70_ner

Sigma-70, non-essential region -

- This hit is scoring at : -122.2
- Scoring matrix: BLOSUM62 (used to infer consensus pattern)

```
112 --- AIESVLQTINPHTGEKVCLSYR--- CADMDREIPALMG--- VSKAVLENVIFV---
Q:
          .:::L...: . E: LS .D D I.. . :: L..
       1 fPqtvdyILaeYdRvetEeqRLsDilsGyiDPddgiapdeAPtathieselaeepssekd
         -----HQDESN----WP1qDPSTLKKKFDDIfsATRYTKALEVIK--
                        ..D:S. P DP...:::F.:: ....K. :.:K
        daadaddddDEdEcceessdddsEagdggp..DPEeArerFgel..reqlektkkalkKh
         ----KLHKDQMQEI----KTFRLKL---ENL-QTVKDQAHKLREN-------
            K :.:.: ...:L : L :.V:...::R:.
        GRgskqalealeaLAelFmpikLvPKQfDaLVervRgmldrvRkqERaIMklCVrdArMP
         ----IAQ------DQEKSDASKS--QMEQLKEKICGTEREILQME
            I.. KS...: .:E::KE.I. .::::..:E
                                                          227
        RkdFiksFpgnETnleWlekllkskkkyadeaLervkedIlrcQqKLadlE
```

Legend of Alignment

- : positive score
- score between -2 and 0

bioSCOUT 1.5.3 - Alignment: 1116E.rad50.sid2 - pfam|hmm|DUF259



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				ģ	Features

Alignment: 1116E.rad50.sid2 - pfam|hmm|DUF259

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|DUF259

Protein of unknown function, DUF259 -

• This hit is scoring at: -137.1

Scoring matrix: BLOSUM62 (used to infer consensus pattern)

Q:	441	DGQIQSKIESMSGILRRRKDKE	KERDAAEVELSK	`	FNLSRID-	
		;;;.::.:.Gt: :			F :::D	
н:	1	adamRamLDqLMGRdasnRngd	esrqkkkvkfdDpeVCrsyLv	rgECPHD)	[FinTKmDn]	LG
		ERERHMQIEVERKTLA	LGERDYDSIISQ	-KRTEVYS	SLEQKIKVL	LR
		: :EK:	_ ERD:.:	:Ē:	L.:KIK	::
		pCpkvHdlklradYErasksrd	yfpkfErdaleflersvsEvt	qsPEiL	ELsEkikEk	πk
		EKDIInrnADERVKLGLKKDAL	ESSKDKLNEIVNEHKDKI	KKVLRGR-		
		É.: D ::KL	E.:::: .: . E.K :	. L .		
		EAfvhDcdrridkakqrL	eetgeeqtkeaaeekRqaeel	laeldee	(AsLPqPvP	Αq
		NPFEK	DMKKEINQAFWPVdKi	EYNELRSI	(SQEAEQ	<u>-</u> E
		PPssELPPPDPRtqEvIgklla	eaEaLGeeGkVdeaqklm.ke	evEeLkal	ckkeleekl	de
		LKFTQSKVTDAreqltklrrdm	dakrrfldsklqsilqisan	vdmfpkVI	LQDAMNKR-	
		::A			D.M.::	
		vrnaapssaqa		Ws	slDeMggqK	Lr
		DEQKRLENFA	NGMREMLAPFEHI	LARK	NHVCPCC	ER
		D:RL.:.	.G :RE.LA:.	K		ER
		VCEvCGAyLsvlDadrRlADHf	gGKlHLGYvkiReklaeLkka	akaklke	evkktgrek	eR
		AFTPDEEDEFVKKQRMQ	717			
		E.:: :. QR				
		-	335			

Legend of Alignment

- positive score
- score between -2 and 0

bioSCOUT 1.5.3 - Alignment: 1116E.rad50.sid2 - pfam|hmm|filament



Projects	Analyses	Admin	Alert	SRS	Tutorial
					Help
				(2) (2)	Features

Alignment: 1116E.rad50.sid2 - pfam|hmm|filament

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|filament

Intermediate filament protein -

This hit is scoring at: -203.9

• Scoring matrix : BLOSUM62 (used to infer consensus pattern)

ο.	940	EEIQ1ELNFLQRTRDTLIVEVDDLRDQHR-MLNEDMSSAQ
Q:	010	E::Q .LN FL::L V::::LR.:
H:	1	nEKeqmQ.nLNDRLAsYIdKVRfLEqqNkeLevkieelrqkqsrggpasvsrlyslYete
		VRWhNAREEKVKASSILERFQKSEEELVLLACE-KEQLIVEKKLL
		:R.: :.:: .R.Q ::.LE: :::. E .L ieeLRrqidqltnerarlqlEidnlrealedfrkKyedKeDLaaQnqlkdlEialntk
		EESLDPLSkEKESLLQEYNALKQKLDEEYHQLAE
٠		EL. E::L ::.LDE. ::LA eaeLaTaL.eRqeaEndlvgLRaQiAklEslaaRkdlDeaTLarvDLEnkvEsLqEElaF
		RKREFQQELDALGRLNMKIKGYLDSKKNEKLKELQgrhvlchSQLQsCM K::E:L :N:::: :Q:: :
		LKknHeEEvkeLqaqiqdtgqvnVEmDaarqqEwklDLtkaLrEiRaQYE.ei
		AKQQRISAELNKSKELLQGQGQLKRNIDDnlkYRKTKADVEQLTRDIESLEERLLSIGSL
		A::.R .AE:L:QRN: .RK.:::L.R.T:SLE .L S: AeknrqeaEewYksKleeLqtaaarngealrsaKeEitElRRqiQsLeiELqslK
		SAIEADLKRHSQEKERLNSEFNRWQGTLSVYQSNISKHKQELKLSQYKDI
	•	S: :: :ER:E. ::Q:S .:: ::E:. L.:Y::: sqnasLErqlaElEeryeaelaqyqalisqlEeeLqqlreEMarqLrEYQeLLdVKlaLD
		EKR 1118 E.R
		iEIATYRKLLEGEESR 359

Legend of Alignment

- positive score
- score between -2 and 0

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Features

Alignment: 1116E.rad50.sid2 - pfam|hmm|Myosin_tail

HMMPFAM - alignment of 1116E.rad50.sid2 against pfam|hmm|Myosin_tail

Myosin tail -

- o This hit is scoring at: -555.1
- Scoring matrix : BLOSUM62 (used to infer consensus pattern)

Q:	237	ASKSQMEQLKEKICGTEREILQMETSLDELRRLQGQIDIKATERS :.Q:L:E:: .E.E: Q:L:: ::.L:.:I :ER:
H:	1	dlerqkreleeqlkrkeselsqlslklEdEqalvaqLqkkikeleaRIeELeEeLEaERa
		TLLTQQHEKLAALSEENEDT
		RIALLETKISKLVRDMDDEASYSSVLSKQNSELTHEIGKLQAEADAHLTMKHERDS ::A.LK:D::K:.S:L. E:L A:.D:K alatLRKKHqdainElsdQieqLqKqKakaEKeKsqlqaEvddllaqldsitKaKlnaEK
		DIKNICTKHNLGPVPehpfTNDVAMNLTNRIKARLSSLENDLLDKKKSNEDQLDVLWKK.: :: .V::L K:RL.S .:DL. : :. E Q:. L K kakqlesQlselqvKldElqRqlnDltsqKsRLqsENsdLtrqleEa£aqvsqlsk
		HYLKINARYSEVDGQIQSKIESMSGILRRRKDKEKERDAAEVELSK ::.: R E :.::::: L. D. E:E.:A .E :LSK lksqlesQLEeAkRslEEEsReRanLqaqlrnlehDlDslrEqlEEEsEAKaeleRqLsk
		FNISRIDERERHMQIEVERKTLALGERDYDSIISQKRTEVYSLEQKIKVLLREKDIIN N :.I.: E : L E : IS: K L :.K. :. an.aeiqqwrsKfEsEgalraEE1EE1KkKlnqkisE1EeaaEaanaKcssLEKtKsRLq
		RNAD~ERVKLGLKKDALESSKDKLNEIVNEHKDKIKKVLRGRNPFEKDMKKEINQAF: ELE::I: E K.K:.:::
		<pre>WpVDKEYNELRSKSQEAEQELKFTQSKVTDAREQLTKLRRDMDAKRRFLDSKLQSIL :EEL:.: ::E K .Q.::.D:QL : R :::RR L:::: r.lKneleElkDqvEaLrRENKnLqdEikDLtdqLgEgGRnvHELEKarRrLEaEkdELq</pre>
		QisanvDmfpkvLQDAMnKRDEQKRLENFANgMREMLAPFE-HLARKnhvcpcceraftp . A : .: A: :::E.K L ::: : E .LA.K aALeEAEaAL.eqeEsKvlRaqvE.lsqiRsEiERRLaEK
		DEEDEFVKKQRMQnsstaerskalAMESSNAEaLFQQLDKLRTIYDAYVKLVEETIP :EE E .:K: A:ES .A. L :. K : K :E .I EEEfEntRKnhqraiesLqas.LaEaEaKgKaEalRlKKKLEgdInELE
		LAEKNLNOHLADESQKAQAFDDLL-GVLAHVQMDRDAVEALLQPTDTIDRHVHEIQQ1vk :A .:.N: A::: :v: :: A E . ::R:Q.

iaLDhaNkanaeaqKnvKkyqqqvkeLQtqvEeeQRaredareqlavaERRataLqa... ${\tt EVEDLEYALDSSGRGVKSLEeiqLELNFLQRTRDTLIVEVDDLRDQHRMLNEDMSSAQVR}$ E:E:L..AL:.: R. K..E .EL :.L.: ..L Q.R.L. ::::.Q ElEELrvaLeqaeRaRKqAE...tElaEaservneLtaqnssLiaqKRKLEgelaalqsD WHNAREEKVKAssilerfQKSEEELVLLAEEkeqLIVEKKLLeESLDPLSKEKESLLQEy ..A .E .A ER :K::.:. LAEE L E:: :.L: L.K: ES ::E LDEavnElkaA...eERakkaqaDaarLaeE...LrqEQehs.qklErlRKqLEsqvKe. nalkQklde-EYHQLAE-------RKREFQQELDALGRLNMKIKGYLdSKKNEKLKE L: :LDE E .L. R RE.:.ELD. R :.:.: L .K...:KE ..LqvRLdEaEaaAlkgGKkvIqKLEaRVReLEaELdgEqRRhaetqKnl.RKaeRrvKE ${\tt LQG------RHVLchsQLQSCMAKQQrisaelNKSKellqqqqqQLKRNIDDNLKYRKTK--}$ LQ ::: :LQ..: K Q K K ..KR.::: :..... LqfQvEEDkKnle...rlQDLvDKLq.....aKiK......tyKRQlEEaEEiaqinl ADVEQLTRDIESLEERL-LSIGSLSAIE---ADLKRH 1075 :...:..R::E..EER. : .SL:.:. A. :R. 864 sKyRkaQreLEdAEERADqAEsslnklRqreaKsRrs

Legend of Alignment

- : positive score
- . score between -2 and 0

SeqWeb Sequence Analysis

HmmerPfam Results

Refine

Query: <u>11168</u>	ID2 from: 1 to: 1316 WPDEF Case 1116 Ra	ad50 SEQ	ID NO:
	quence family classification (score includ	des all	
domains):			-
Model	Description	Score	F:-
value N			
	Rad50 zinc hook motif	36.3	6.9e-
HSP9 HSP12	Heat shock protein 9 / 12	-11.4	
4.1 1			
LEA 1	Late embryogenesis abundant (LEA) group	-14.3	
5.6 1			
	H-NS histone family	-28.3	
7.9 1			
ABC tran	ABC transporter	-46.1	
0.14 1			
SMC_N	RecF/RecN/SMC N terminal domain	-69.5	
0.75 1		·	
TACC	Transforming acidic coiled-coil-contain	-	
88.9	8 1		
DUF164	Uncharacterized ACR, COG1579	-90.1	
4.1 1			
Tropomyosin	Tropomyosin	_	
91.4	8 1.		
SMC_C	SMC family, C-terminal domain	-116.7	
0.36 1			
<u>sigma70_ner</u>	Sigma-70, non-essential region	-122.2	
8.3 1			
<u>filament</u>	Intermediate filament protein	-203.8	
3.3 1			

SeqWeb Sequence Analysis

ERM Ezrin/radixin/moesin family -236.2

9.5 1

Parsed for domains:

Alignments of top-scoring domains:

Rad50_zn_hook: domain 2 of 2, from 673 to 726: score 32.5, E = 9.9e-06
*->galesekaelkkaieeleeeesscCPvCgReLgteeekkelikeyks

+ ++++++1 ++ ++ ++ CP+C+R + t++e+ e++k+++

1116SID2 673 NFANGMREMLAPFEHLARKNHV--CPCCERAF-TPDEEDEFVKKQRM
716

eldrlpeelk<-* + +++ e k 1116SID2 717 QNSSTAERSK 726

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g die	Accession number: PF04423
. S.	Rad50 zinc hook motif
	The Mre11 complex (Mre11 Rad50 Nbs1) is central to chromosomal maintenance and functions in homologous recombination, telomere maintenance and sister chromatid association. The Rad50 coiled-coil region contains a dimer interface at the apex of the coiled coils in which pairs of conserved Cys-X-X-Cys motifs form interfocking hooks that bind one Zn ion. This alignment includes the zinc hook motif and a short stretch of coiled-coil on either side.
	INTERPRO description (entry IPR007517)
Figure 1: 118d Replication Rad50 coiled-coil zn hook	The Mre11 complex (Mre11 Rad50 Mbs1) is central to chromosomal maintenance and functions in homologous recombination, telomere maintenance and sister chromatid association. The Rad50 colled-coil region contains a dimer interface at the apex of the colled coils in which pairs of conserved Cys-X-X-Cys motifs form interlocking hooks that bind one Zn ion. This alignment includes the zinc hook motif and a short stretch of colled-coil on either side.
Key:	
Domain Chain Residue Residue	
Rad50 zn hook A 421 475	
Rad50 zn hook B 421 475	
The Swissprot/PDB mapping was provided by MSD	

Alignment Domain organisation

(17) O Ful	As a Graphic As a Tree Zoom 0.5 pixels/aa.	here	Phylogenetic tree	© Seed (17) O Full (25)	Download free ATV Applet	The trees were generated using <u>Quicktree</u> To find out more about ATV phylogenetic tree-viewer <u>click</u> <u>here</u>
© Seed (17) O Full (25)	Format Coloured alignment Get alignment View HMM logo	Further alignment options <u>here</u> Help relating to Pfam alignments <u>here</u>	Species Distribution		NEW! View alignments & domain organisation by species	View Species Tree

Pram: Rad50_zn_book

		Database References		
PDB You can find out how to set up Rasmol here	118d A; 421; 475;	CONTRIBUTION (CONTRIBUTION)	Scotleanfraur Westernamen	
SYSTERS	Rad50 zn hook			
PANDIT	Rad50 zn hook			

Pfam: Rod50_zn_book

		The state of the same of the s
Literature References	Pfam specific information	
1.	Author of entry	Bateman A
Tethering on the brink: the evolutionarily conserved Mre11-Rad50 complex.	Type definition	Motif
Connelly JC, Leach DR; Trands Blochem Sci 2002;27:410-418.	Alignment method of seed	Clustalw
2.	Source of seed members	Bateman A
The Rad50 zinc-hook is a structure joining Mre11 complexes in DNA	Average Length	55.6
Hopfing KP, Chaig L, Moncallan G, Zinkel RA, Usul T, Owen BA, Karcher A, Henderson B, Racher H. McManzav Co. Carney 3P. Petriki JH. Tainer 74.	Average %id	24
Nature 2002;418:562-566.	Average Coverage	5,34%

	HMMER build information	ion
	Pfam_Is [Download HMM]	Pfam_fs [Download HMM]
Gathering cutoff	25.0 25.0;	25.0 25.0
Trusted cutoff	40.9 40.9;	31.4 38.9
Noise cutoff	22.6 22.6;	24.6 20.6
Build method of HMM	Build method of HMM hmmbuild -F HMM Is SEED hmmcalibrateseed 0 HMM Is	hmmbuild -F HMM_Is SEED hmmbuild -f -F HMM_Is SEED hmmcalibrateseed 0 HMM_Is hmmcalibrateseed 0 HMM_fs

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Pfam: Mre11_DNA_bind



Protein families database of alignments and HMMs



Keyword Search Protein Search | Browse Pfam Mre11 DNA bind

DNA Search

Accession number: PF04152

Previous Identifiers: Mer11_DNA_blnd;

Mre11 DNA-binding presumed domain



The Mre11 complex is a multi-subunit nuclease that is composed of Mre11, Rad50 and Nbs1/Xrs2, and is involved in checkpoint signalling and DNA replication [1]. Mre11 has an intrinsic DNA-binding activity that is stimulated by Rad50 on its own or in combination with Nbs1 [2].

INTERPRO description (entry IPR007281)

The Mre11 complex is a multi-subunit nuclease that is composed of Mre11, Rad50 and Nbs1/Xrs2, and is involved in checkpoint signalling and DNA replication [MEDLINE:21984524]. Mre11 has an intrinsic DNA-binding activity that is stimulated by Rad50 on its own or in combination with Nbs1 [MEDLINE:20300914].

Alignment	Domain organisation	
● Seed (10) ○ Full (20)	● Seed (10) ○ Full (20)	
Format Coloured alignment Get alignment View HMM logo Further alignment options here Help relating to Pfam alignments here	As a Graphic As a Tree Zoom 0.5 pixels/aa Bootstrap tree View Graphic NIFAS Applet To find out about the NIFAS tree-viewer, dick here	
Species Distribution	Phylogenetic tree	
NET! View alignments & domain organisation by	● Seed (10) ○ Fuil (20)	
species Tree depth : Show all levels View Species Tree	The trees were generated using <u>Quicktree</u> To find out more about ATV phylogenetic tree-viewer <u>click</u> <u>here</u>	

	Patabase References
Ver The Control of th	Mre11 DNA bind

Plam: Mrc11_DNA_bind

Mre11_DNA bind **PANDIT**

Literature References	
1. A mechanistic basis for Mre11-directed DN/ Joining at microhomologies.	<u>.</u>
Pauli TT, Gellert M; Proc Nati Acad Sci U S A 2000;97:6409-6414.	
2.	
The Mre11 complex: at the crossroads of	
dna repair and checkpoint signalling.	
D'Amours D, Jackson SP; Nat Rev Mol Celt Blol 2002;3:317-327-	

Pfam specific information		
Author of entry Wood V, Finn RD		
Type definition	Domain	
Alignment method of seed Clustalw		
Source of seed members Pfam-B_3909 (release 7.3);		
Average Length	200.3	
Average %id	39	
Average Coverage	28.59%	

HMMER build information				
	Pfam_Is [Download HMM]	Pfam_fs [Download HMM]		
Gathering cutoff	25.0 25.0;	25.0 25.0		
Trusted cutoff	71.9 71.9;	49.5 32.0		
Noise cutoff	13.1 13.1;	8.2 17.4		
Build method of HMM	hmmbuild -F HMM_ls SEED hmmcalibrate seed 0 HMM_ls	hmmbuild -f -F HMM_fs SEED hmmcalibrateseed 0 HMM_fs		

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